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54 **Caries-preventive composition.**

57 A caries-preventive composition comprises an antibody obtained by immunizing a mammal with at least one antigen selected from the group consisting of Streptococcus mutans, its cell-wall fraction, fibrous substance fraction, glucosyl-transferase fraction and protein antigen fraction, and a synergist selected from the group consisting of fluorine compounds, chlorhexidine and its salts, lytic enzymes, bacteriocins, glucosyltransferase inhibitors, proteases and dextranases.

"CARIES-PREVENTIVE COMPOSITION"

This invention relates to a caries-preventive composition which, when applied to the mouth, can prevent dental caries by suppressing formation of dental plaque.

5 Dental plaque firmly adhering to the surface of teeth is composed of about 70% bacteria, about 20% polysaccharides produced by the bacteria and about 10% food remains. It is said that acids stored in dental plaque decalcify enamel, causing dental caries. There-
10 fore, dental plaque is observed as a cause of dental caries.

Formation of dental plaque is accelerated due to the synthesis of polysaccharides from sucrose by oral bacteria, especially *Streptococcus mutans*. In
15 more detail, *Streptococcus mutans* synthesizes adhesive polysaccharides such as dextran and mutan from sucrose through the production of GTF (glucosyltransferase, dextran-synthesizing enzyme). The thus synthesized polysaccharides incorporate *Streptococcus mutans* as
20 well as other bacteria (viruses), forming dental plaque having a given bacterial bouquet. In addition, bacteria such as *Streptococcus mutans* produce acids by utilizing various kinds of sugar and the thus produced acids decalcify the surface of enamel by remaining in poly-
25 saccharides and bacterial walls.

Accordingly, it is desirable to decrease the number of Streptococcus mutans in the mouth and suppress the formation of dental plaque in order to prevent dental caries.

5 It is known in British patent No.1,505,513 that colonization of Streptococcus mutans in the mouth is suppressed by using mother's milk obtained by immunizing a cow with whole bacterial bodies of Streptococcus mutans.

10 The present inventors studied antibodies which are amongst the antibodies to various antigens derived from Streptococcus mutans and inhibit the colonization of Streptococcus mutans in the mouth. As a result, the inventors found that antibodies contained in anti-
15 serum and milk obtained by immunizing mammals with Streptococcus mutans, its cell-wall fraction, fibrous substance fraction, glucosyltransferase fraction and protein antigen fraction have certain degrees of dental-plaque-formation suppressing effect. However, the
20 effect was not necessarily sufficient and a higher effect of suppressing the formation of dental plaque was necessary.

 An object of the present invention is to provide a caries-preventive composition having an excellent
25 effect in preventing dental caries.

 For the purpose of attaining the above object, the present inventors further conducted an intensive study, and, as a result, found that the combination

of such an antibody and a fluorine compound, chlorhexidine or a chlorhexidine salt, a lytic enzyme, a bacteriocin, a glucosyltransferase inhibitor, a protease or a dextranase works effectively for the prevention of dental
5 caries by causing a significantly increased dental-plaque-formation suppressing effect through the suppression of colonization of *Streptococcus mutans*.

Therefore, this invention provides a caries-preventive composition characterized by being composed
10 of the combination of antibody obtained by immunizing a mammal with at least one antigen selected from the group consisting of *Streptococcus mutans*, its cell-wall fraction, fibrous substance fraction, glucosyltransferase fraction and protein antigen fraction with at least one
15 synergist selected from the group consisting of fluorine compounds, chlorhexidine and its salts, lytic enzymes, bacteriocins, glucosyltransferase inhibitors, proteases and dextranases.

According to this invention, since the combination
20 of said antibody and said synergist component exerts a synergistic effect on the inhibition of colonization of *Streptococcus mutans* in the mouth, the formation of dental plaque is efficiently suppressed, resulting in the effective prevention of dental caries.

25 In addition, since said antibody and said synergist component both are quite safe, the caries-preventive composition of this invention can be safely used.

The above and other objects, features, and advantages of this invention will be more fully understood by reading the following description.

The caries-preventive composition according to this invention is prepared by use of antibody contained in antiserum and/or milk obtained by immunizing a mammal with at least one antigen selected from the group consisting of *Streptococcus mutans*, its cell-wall fraction, fibrous substance fraction, glucosyltransferase (GTF) fraction and protein antigen fraction as described above. It should be noted that the fibrous substance means a pili-like or fimbriae fraction.

Streptococcus mutans used as an antigen may be prepared through well-known culture and pretreatment carried out by, for example, growing bacteria in external solution obtained by the dialysis of BHI medium before the thus grown bacteria are washed and subjected to formalin treatment. *Streptococcus mutans* separated from human mouth and belonging to the serotypes C, D, E, F and G may preferably be used, particularly one belonging to the serotype-C which is numerous in the human mouth. Such *Streptococcus mutans* includes NCTC10449, Ingbritt, OMZ70, JC-2, etc. and their mutant strains.

The cell-wall fraction of *Streptococcus mutans* may be prepared, for example, according to the method of Bleiweis et al. (*J. Bacteriol.*, 88, 1198-1200, 1964) by subjecting *Streptococcus mutans* to crushing treatment in a Brown's cell crusher and glass beads of 0.17 to

0.18 mm diameter, then treating the thus obtained cell walls with trypsin to remove protein contaminating the cell walls, followed by washing the cell walls with distilled water before they are lyophilized. The fibrous (pili-like or fimbriae) substance fraction may be prepared, for example, according to the method of J. Van Hoate et al. (Arch. Oral. Bio., 16, 1131-1141, 1971) by culturing Streptococcus mutans in a medium obtained by the dialysis of BHI medium and containing 5% sucrose under an anaerobic condition, then centrifuging the culture medium to obtain a supernatant solution, then adding three times as much ethanol as the supernatant solution by volume, followed by collecting the precipitate of the thus obtained solution. As the fibrous substance fraction there may also be used a pili-like structure from the cell wall of Streptococcus mutans and its purified substance prepared by the ordinary cell wall extract method from the cultured bacteria, using solvents such as phosphate buffer containing 1M sodium chloride according to the method of Tsurumizu et al. (Japanese Journal of Bacteriology, 38, (1) 471, 1983). The GTF fraction may be prepared, for example, according to the method of Inoue et al. (Microbial Aspects of dental caries Vol. III, 665-682, 1976 [Information Retrieval Inc.]) using a solution prepared by the following method: after Streptococcus mutans is implanted and grown in a medium obtained by the dialysis of BHI medium, the bacterial bodies are removed by centrifugation and the supernatant is saturated with ammonium sulfate at the level

of 40%, followed by dialyzing the precipitate of the 40% ammonium sulfate fraction against 50 mM phosphate buffer solution and concentrating or diluting the obtained solution. The protein antigen fraction may be prepared, for example, according to the method of Lehner et al. (J. General Microbiology, 122, 217-225, 1981) by culturing Streptococcus mutans in a medium obtained by the dialysis of BHI medium, then centrifuging the culture medium to obtain a supernatant solution, followed by fractionation with 75% ammonium sulfate solution to collect the precipitate; the thus obtained precipitate is then subjected to DE-52 column chromatography under the existence of 6M urea, and the protein antigen fraction is dissolved in physiological saline, this being followed by dialyzing the thus obtained solution whereafter the dialyzed solution is subjected to gel filtration through Sepharose CL6B.

The usual method may be adopted in immunizing mammals with said antigens. As mammals to be immunized, goats, sheep, horses, cows or rabbits may be used.

The antibody (protein fraction in the antiserum and the milk) may be separated from the antiserum and the milk according to the ordinary antibody purification method including the salting-out method, the gel-filtration method, ion-exchange chromatography or affinity chromatography, the salting-out method using ammonium sulfate being preferred. In the salting-out method, the antiserum or the milk is saturated with ammonium sulfate, preferably at the level of not more than 40%, to produce

the precipitate, followed by dialyzing the precipitate against physiological saline to obtain the purified precipitate as the antibody. The preferred antibody is obtained from the equine antiserum and the bovine antiserum and
5 milk.

In this invention, the antibody contained in the antiserum and milk obtained by immunizing the mammal with said antigen is blended into the composition. In this case, the antiserum and milk as well as the antibody separated and purified therefrom may be used. Each of these materials may be used alone or in a combination of two or more.

The caries-preventive composition according to this invention is prepared by the combination of said
15 antibody and at least one synergist selected from the group consisting of fluorine compounds, chlorhexidine and its salts, lytic enzymes, bacteriocins, glucosyltransferase inhibitors, proteases and dextranases.

As fluorine compounds, sodium fluoride, potassium
20 fluoride, lithium fluoride, ammonium fluoride, sodium monofluorophosphate, sodium hydrogen monofluorophosphate, potassium monofluorophosphate, ammonium monofluorophosphate, potassium hexafluorozirconate, and potassium hexafluorotitanate may be used. Also useful are cesium fluoride,
25 nickel fluoride, zirconium fluoride, silver fluoride, hexylamine hydrofluoride, laurylamine hydrofluoride, cetylamine hydrofluoride, glycine hydrofluoride, lysine hydrofluoride, alanine hydrofluoride and the like. Among

them, monofluorophosphates such as sodium monofluorophosphate and potassium monofluorophosphate, alkali-metal fluorides such as sodium fluoride, potassium fluoride and ammonium fluoride, fluorides containing stannous tin
5 such as stannous fluoride and stannous chloride fluoride and the like may preferably be used. Especially, sodium monofluorophosphate, sodium fluoride and stannous fluoride are more preferably used.

As chlorhexidine salts, chlorhexidine hydrochloride
10 or chlorhexidine gluconate can be used.

As lytic enzymes, those derived from *Streptomyces griseus*, *Streptomyces diastatochromagenes*, *Streptomyces farinosus*, *Chalaropsis*, *Flavobacterium*, *Myxobacter*, *Staphylococcus epidermidis*, *Micrococcus*, *Pseudomonas aeruginosa*,
15 *Aeromonas*, *Streptomyces albus* and *Streptomyces globisporus* can be used.

As bacteriocins, those derived from *Enterobacter cloacae*, *Escherichia coli*, *Proteus mirabilis*, *Pseudomonas aeruginosa*, *Streptococcus mutans* and *Staphylococcus*
20 *staphylococcus* can be used.

As GTF inhibitors, those derived from *Arthrrium* sp., *Fusarium* sp., *Macrophomina* sp., *Micromonospora* sp., *Gnomoniella* sp., *Nodulisporium* sp., and *Aspergillus* sp., can be used, and more specifically, those described in
25 Japanese Patent Application Laid-Open Nos.56-103193, 57-28097, 57-98215 and 57-146587 can be used.

As proteases, those derived from *Aspergillus* sp., and *Bacillus* sp., can be used.

As dextranases, those derived from *Chaetomium* sp., *Streptomyces* sp., *Bacillus* sp. and *Corynebacterium* can be used.

In this invention, each of these synergist components may be used alone or in a combination of one or two.

The caries-preventive composition according to this invention can be prepared and used in various forms applicable to the mouth such as dentifrices (including toothpaste, toothpowder and liquid dentifrice), mouthwashes, dental pastes, gingival massage creams, gargle tablets, troches, chewing gums, ice-creams, whipped creams and the like.

The antibody and the synergist component may be mixed in a given form. Alternatively, the antibody and the synergist component may be jointly used after they are prepared separately.

It is preferred that the quantity of said antibody administered is 0.0001 to 50 g/kg/day. As to the quantity of said synergist component administered, a quantity corresponding to 0.0001 to 1 g/kg/day fluorine for fluorine compounds, a quantity corresponding to 0.0001 to 1 g/kg/day chlorhexidine for chlorhexidine and its salts, a quantity

of 0.0001 to 10 g/kg/day each for lytic enzymes, bacteriocins and glucosyltransferase inhibitors and a quantity of 0.0001 to 5 g/kg/day each for proteases and dextranases are preferably used. The blended amount of the antibody to the oral composition may be in the range of 0.0002 to 10%, preferably 0.002 to 5% by weight of the total weight of the composition. As to the blended amount of the synergist component in the composition, it is preferred that an amount corresponding to 0.0001 to 0.1 wt%, preferably 0.0001 to 0.001 wt% fluorine for fluorine compounds; an amount corresponding to 0.1 to 1000 ppm, preferably 10 to 100 ppm chlorhexidine for chlorhexidine and its salts; and an amount of 0.0001 to 10 wt%, preferably 0.001 to 5 wt% each for lytic enzymes, bacteriocins, glucosyltransferase inhibitors, proteases and dextranases may be blended to the composition.

The oral composition according to this invention may further include additional well-known ingredients depending on the type and form of a particular oral composition. Any desired known ingredients may be mixed with said antibody and synergist component.

In preparing dentifrice compositions, an abrasive may be blended generally in an amount of 5 to 95%, especially 15 to 60% by weight of the composition, including calcium secondary phosphate dihydrate, calcium secondary phosphate anhydrate, calcium primary phosphate, calcium tertiary phosphate, calcium carbonate, calcium

pyrophosphate, insoluble sodium metaphosphate, magnesium **0140498**
tertiary phosphate, magnesium carbonate, calcium sulfate,
titanium dioxide, resins, and the like.

In preparing paste-like compositions, typically
5 toothpastes, a binder may be blended generally in an amount
of 0.3 to 5% by weight, including sodium carboxymethyl
cellulose, methyl cellulose, sodium carboxymethyl
hydroxyethyl cellulose, hydroxyethyl cellulose, gum arabic,
tragacanth gum, karaya gum, polyvinylalcohol, sodium
10 polyacrylate, carboxyvinyl polymer, polyvinyl pyrrolidone,
and the like.

In preparing paste-like and liquid oral
compositions, typically toothpastes and mouthwashes, a
humectant may be blended generally in an amount of 10 to
15 70% by weight, including polyethylene glycol, ethylene
glycol, sorbitol, glycerol, propylene glycol, 1,3-butylene
glycol, xylitol, maltitol, lactitol, and the like.

In addition to the above ingredients, a surface
active agent including water soluble salts of alkyl sulfate
20 having 8 to 18 carbon atoms such as sodium laurate and
sodium myristate, sodium salts of higher fatty acids,
water-soluble salts of sulfonated monoglycerides of higher
fatty acids having 10 to 18 carbon atoms in the fatty acid
group such as sodium lauryl monoglyceride sulfonate and
25 sodium coconut monoglyceride sulfonate, sodium monoglyceride
monosulfates of higher fatty acids, olefin sulfonates,
paraffin sulfonates, sodium N-methyl-N-palmitoyl touride,

sodium N-lauroyl sarcosinate, sodium N-lauroyl-β-alanine, stearyl monoglyceride, sucrose fatty acid esters having 12 to 18 carbon atoms in the fatty acid group such as sucrose monolaurate and dilaurate, lactose fatty acid esters, lactitol fatty acid esters, maltitol fatty acid esters, stearic acid monoglyceride, polyoxyethylene sorbitan monolaurate, polyoxyethylene-hardened castor oil, condensates of sorbitan monostearate with approximately 60 moles of ethylene glycol, condensates of ethylene oxide with propylene oxide, and their derivatives such as polyoxyethylene polyoxypropylene monolauryl ester, betaine and amino acid type amphoteric surfactants, and the like may be blended in an amount of 0 to 10%, preferably 0.1 to 5%, more preferably 1 to 2.5% by weight of the composition. A flavor such as an essential oil including peppermint oil and spearmint oil and a flavoring material including l-menthol, carvone, eugenol and anethole, a sweetener such as sodium saccharinate, stevioside, neohesperidyl dihydrochalcone, glycyrrhizin, perillartine, p-methoxycinnamic aldehyde, a preservative, and the like may be blended in an effective amount.

In this invention, effective ingredients such as mutanase, sorbic acid, alexidine, hinokitiol, cetylpyridinium chloride, alkyl glycine, alkyl diaminoethyl glycinate, allantoin, ε-aminocaproic acid, tranexamic acid, azulene, vitamin E, a water soluble primary or secondary phosphate, a quaternary ammonium compound, sodium chloride

and crude drugs may also be blended in an effective amount.

Other types of compositions may also be prepared by selecting any desired ingredients as usual and mixing them by a conventional procedure.

5 Examples of the other ingredients for various types or forms of the composition are shown in the following Examples.

 Paste-like and liquid oral compositions may generally have a pH ranging from 5 to 10, but not limited
10 thereto.

 The caries-preventive composition according to this invention, owing to the combination of said antibody and said synergist component, can efficiently suppress the formation of plaque caused by Streptococcus mutans, thereby
15 excellently preventing the formation of dental caries.

 Examples of this invention will be given in the following although this invention is not restricted to them.

EXAMPLE 1

 Antisera and mother's milks were obtained by using
20 the following antigens according to the following method.

(1) Antigens

Streptococcus mutans NCTC10449

 Bacteria grown in the external solution obtained by the dialysis of BHI medium, after being washed, were
25 treated with formalin before being supplied for use.

Cell-wall fraction of Streptococcus mutans NCIC10449

The fraction prepared according to the method of Bleiweis et al. (J. Bacteriol., 88, 1198-1200, 1964) was supplied for use.

5 Fibrous substance fraction of Streptococcus mutans
 NCIC10449

The fraction prepared according to the method of J. Van Hoate et al. (Arch. Oral. Bio., 16, 1131-1141, 1971) and Tsurumizu et al (Jap. J. Bacteriology, 38,
10 (1) 471, 1983) were supplied for use.

Glucosyltransferase fraction of Streptococcus mutans
NCIC10449

The fraction prepared according to the method of Inoue et al. (Microbial Aspects of dental caries Vol.
15 III, 665-682, 1976 [Information Retrieval Inc.]) was
supplied for use.

Protein antigen fraction of Streptococcus mutans
NCIC10449

The fraction prepared according to the method
20 of Lehner et al. (J. General Microbiology, 122, 217-225,
1981) was supplied for use.

(2) Preparation of Antiserum and Mother's Milk

Said antigen was mixed with Freund's complete
adjuvant, and a pregnant goat, horse, cow or rabbit was
25 immunized with the thus prepared mixture. After the animal
was immunized three times with the mixture of said antigen
and Freund's incomplete adjuvant before its delivery, the

colostrum was collected after the delivery. As to the antiserum, after the animal was immunized four times in the same manner as above, the blood was collected and coagulated, and supernatant solution obtained by
5 centrifuging the coagulated blood was used as a sample.

An antibody is prepared by adding ammonium sulfate to the antiserum to saturate it at the level of 40%, separating the obtained precipitate by centrifugation, and dialyzing the precipitate against physiological saline,
10 and the inner solution was used as a sample.

Next, the colonizing tests of Streptococcus mutans in the mouth were conducted according to the following method by using said antiserum and mother's milk as well as a fluorine compound, a chlorhexidine salt, a lytic
15 enzyme, a bacteriocin, GTF inhibitors, a protease and a dextranase used as synergist components.

(3) Colonization of Streptococcus mutans in the Mouth

After male hamsters of five week old were divided into groups each consisting of five individuals, each
20 hamster was inoculated with 1×10^8 bacteria of Streptococcus mutans of the NCTC10449 strain. From the day of the inoculation, drinking water containing the effective components (said antiserum or milk and the synergist component) was administered to each hamster. One week and
25 four weeks after the start of the administration, the teeth of each hamster were rubbed with a cotton ball before it is immersed in a small amount of physiological saline to

disperse bacteria homogeneously in it. After a given amount of the thus obtained solution was scattered on the BHI plate medium and the mitis salivalius plate medium, the number of whole bacteria and the number of the colonies of

5 Streptococcus mutans were counted. The number of Streptococcus mutans was indicated by the number of Streptococcus mutans per 10,000 whole bacteria. The concentration of antiserum or mother's milk in the drinking water was adjusted to 0.025%. As to the concentration of

10 the synergist component, it was adjusted to 0.05% for a fluorine compound (NaF), 0.005% for a chlorhexidine salt (chlorhexidine gluconate), 0.05% for a lytic enzyme, 0.01% each for a bacteriocin, a GTF inhibitor and a protease and 0.005% for a dextranase.

15 For comparison, the same experiments were conducted without jointly using antiserum or mother's milk and the synergist component by adding antiserum or mother's milk alone, by adding the synergist component alone and by adding none of antiserum, mother's milk and the synergist

20 component (Control).

The results obtained by using the fluorine compound (NaF) as the synergist component are indicated in Table 1; those obtained by using chlorhexidine gluconate (CHX), in Table 2; those obtained by using the lytic enzyme,

25 in Table 3; those obtained by using the bacteriocin, in Table 4; those obtained by using the GTF inhibitors, in Table 5; those obtained by using the protease, in Table

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6; and those obtained by using the dextranase, in Table 7.

Table 1

| Samples Added | Number of S. mutans bacteria 1 week after | Number of S. mutans bacteria 4 weeks after |
|--|--|---|
| Control | 3890 | 4250 |
| Goat anti-whole- bacteria serum | 2178 | 1467 |
| " + NaF | 1945 | 297 |
| Goat anti-GTF serum | 1828 | 1510 |
| " + NaF | 1556 | 212 |
| Goat anti-whole- bacteria mother's milk | 1984 | 1382 |
| " + NaF | 1945 | 170 |
| Goat anti-cell- wall serum | 1750 | 1340 |
| " + NaF | 1750 | 255 |
| Goat anti-protein serum | 1984 | 1255 |
| " + NaF | 1945 | 85 |
| Goat anti-fibrous- substance milk | 1945 | 1340 |
| " + NaF | 1711 | 127 |
| NaF alone | 3112 | 2040 |

Table 2

| Samples Added | Number of S. mutans bacteria 1 week after | Number of S. mutans bacteria 4 weeks after |
|--|--|---|
| Control | 3890 | 4250 |
| Antibody from equine anti-whole-bacteria serum | 2139 | 1425 |
| " + CHX | 2139 | 212 |
| Antibody from equine anti-GTF serum | 1789 | 1297 |
| " + CHX | 1634 | 340 |
| Equine anti-whole- bacteria mother's milk | 1945 | 1340 |
| " + CHX | 1984 | 170 |
| Equine anti-cell- wall serum | 2022 | 1425 |
| " + CHX | 1867 | 297 |
| Equine anti-protein serum | 2023 | 1425 |
| " + CHX | 1945 | 212 |
| Equine anti-fibrous- substance milk | 1867 | 1383 |
| " + CHX | 1134 | 85 |
| CHX alone | 3112 | 2975 |

| Samples Added | Number of S. mutans bacteria 1 week after | Number of S. mutans bacteria 4 weeks after |
|---|--|---|
| Control | 3890 | 4250 |
| Bovine anti-whole- bacteria serum | 2178 | 1425 |
| " + Lytic enzyme | 1945 | 255 |
| Bovine anti-GTF serum | 1828 | 1382 |
| " + Lytic enzyme | 1556 | 340 |
| Bovine anti-whole- bacteria mother's milk | 2023 | 1298 |
| " + Lytic enzyme | 1945 | 127 |
| Antibody from bovine anti-cell-wall serum | 2139 | 1255 |
| " + Lytic enzyme | 1867 | 85 |
| Bovine anti- protein serum | 1556 | 1085 |
| " + Lytic enzyme | 1556 | 42 |
| Bovine anti-fibrous substance milk | 2334 | 1298 |
| " + Lytic enzyme | 1984 | 85 |
| Lytic enzyme alone | 3112 | 3485 |

Note) As the lytic enzyme, one obtained from Streptomyces globisporus was used.

Table 4

| Samples Added | Number of S. mutans bacteria 1 week after | Number of S. mutans bacteria 4 weeks after |
|--|--|---|
| Control | 3890 | 4250 |
| Rabbit anti-whole- bacteria serum | 2178 | 1383 |
| " + Bacte- riocin | 2100 | 298 |
| Rabbit anti-GTF serum | 1556 | 1467 |
| " + Bacte- riocin | 1634 | 255 |
| Rabbit anti-whole- bacteria mother's milk | 1945 | 1510 |
| " + Bacte- riocin | 1945 | 213 |
| Rabbit anti-cell- wall serum | 2023 | 1595 |
| " + Bacte- riocin | 1634 | 170 |
| Rabbit anti-protein serum | 1945 | 1298 |
| " + Bacte- riocin | 1751 | 128 |
| Rabbit anti-fibrous- substance milk | 2178 | 1383 |
| " + Bacte- riocin | 1751 | 128 |
| Bacteriocin alone | 2723 | 3060 |

Note) As the bacteriocin, one obtained from Streptococcus L-1, microbial technology research laboratory trust number 3220, was used.

Table 5

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| Samples Added | Number of S. mutans bacteria 1 week after | Number of S. mutans bacteria 4 weeks after |
|---|--|---|
| Control | 3890 | 4250 |
| Goat anti-whole- bacteria serum | 2188 | 1425 |
| " + GTF in- hibitor A | 2100 | 212 |
| Goat anti-GTF serum | 1867 | 1297 |
| " + GTF in- hibitor A | 1789 | 176 |
| Goat anti-whole- bacteria mother's milk | 1945 | 1340 |
| " + GTF in- hibitor B | 1828 | 85 |
| Goat anti-cell- wall serum | 2022 | 1425 |
| " + GTF in- hibitor A | 1945 | 340 |
| Goat anti- protein serum | 1945 | 1297 |
| " + GTF in- hibitor C | 1906 | 255 |
| Goat anti-fibrous substance milk | 2178 | 1383 |
| " + GTF in- hibitor A | 1751 | 128 |
| GTF inhibitor A alone | 2723 | 3485 |
| GTF inhibitor B alone | 3112 | 3400 |
| GTF inhibitor C alone | 2995 | 3315 |

Note) GTF inhibitor A was obtained from *Aspergillus terreus*;
GTF inhibitor B, from *Arthrinum* sp. M 5071; and
GTF inhibitor C, from *Micromonospora* sp. SF-2259.

Table 6

| Samples Added | Number of S. mutans bacteria 1 week after | Number of S. mutans bacteria 4 weeks after |
|--|--|---|
| Control | 3890 | 4250 |
| Equine anti-whole- bacteria serum | 2334 | 1297 |
| " + Protease | 2022 | 85 |
| Equine anti-GTF serum | 1867 | 1340 |
| " + Protease | 1789 | 85 |
| Equine anti-whole- bacteria mother's milk | 2022 | 1382 |
| " + Protease | 1867 | 42 |
| Equine anti-cell- wall serum | 2178 | 1510 |
| " + Protease | 2022 | 170 |
| Equine anti-protein serum | 1945 | 1552 |
| " + Protease | 1711 | 42 |
| Equine anti-fibrous- substance milk | 2100 | 1340 |
| " + Protease | 1634 | 170 |
| Protease alone | 3034 | 3655 |

Note) The protease used is derived from Aspergillus sp.

Table 7

| Samples Added | Number of S. mutans bacteria 1 week after | Number of S. mutans bacteria 4 weeks after |
|---|--|---|
| Control | 3890 | 4250 |
| Bovine anti-whole- bacteria serum | 2334 | 1297 |
| " + Dextra- nase | 2100 | 212 |
| Bovine anti-GTF serum | 2022 | 1510 |
| " + Dextra- nase | 1789 | 212 |
| Bovine anti-whole- bacteria mother's milk | 2139 | 1552 |
| " + Dextra- nase | 1945 | 340 |
| Bovine anti-cell- wall serum | 2139 | 1595 |
| " + Dextra- nase | 1828 | 85 |
| Bovine anti- protein serum | 2022 | 1297 |
| " + Dextra- nase | 1983 | 170 |
| Bovine anti-fibrous substance milk | 1867 | 1383 |
| " + Dextra- nase | 1634 | 85 |
| Dextranase alone | 2022 | 1275 |

Note) The dextranase used is derived from Chetomium sp.

From the results indicated in Tables 1 to 7, it is found that the combination of the antiserum or mother's milk and the synergist component according to this invention excellently suppresses the colonization of Streptococcus mutans.

EXAMPLE 2 Toothpaste

| | | |
|----|---------------------------------------|---------|
| | Calcium secondary phosphate dihydrate | 50.0% |
| | Glycerol | 20.0 |
| | Sodium carboxymethylcellulose | 1.0 |
| 10 | Sodium lauryl sulfate | 1.5 |
| | Sodium lauroyl sarcosinate | 0.5 |
| | Flavor | 1.0 |
| | Sodium saccharinate | 0.1 |
| | Water | Balance |
| 15 | | 100.0% |

The above components were blended with 0.1% or 0.2% antibody of goat whole-bacteria and 0.1% sodium fluoride, 0.01% chlorhexidine gluconate, 0.1% a lytic enzyme, 0.01% a bacteriocin, 0.001% a protease, 0.1% GTF inhibitor-A or 0.25% (3000 units/g) a dextranase.

EXAMPLE 3 Toothpaste

| | | |
|----|-------------------------------|-------|
| | Calcium secondary phosphate | 50.0% |
| | Sorbitol | 10.0 |
| | Glycerol | 10.0 |
| 25 | Sodium carboxymethylcellulose | 1.0 |
| | Sodium lauryl sulfate | 2.0 |
| | Flavor | 1.0 |

| | | |
|---------------------|---------|---------|
| Sodium saccharinate | 0.1 | 0140498 |
| Ethanol | 2.0 | |
| Mutanase | 0.1 | |
| Water | Balance | |
| | | <hr/> |
| | 100.0% | |

5

The above components were blended with 0.1% bovine anti-cell-wall serum and 0.3% sodium monofluorophosphate, 0.01% chlorhexidine gluconate, 0.05% a lytic enzyme, 0.02% a bacteriocin, 0.001% a protease, 0.1% GTF inhibitor-C or 0.25% a dextranase.

10

EXAMPLE 4 Toothpaste

| | |
|--|----------------|
| Calcium carbonate | 50.0% |
| Glycerol | 20.0 |
| Sodium carboxymethylcellulose | 1.5 |
| Sodium carboxymethylcellulose | 1.0 |
| Sodium lauryl sulfate | 0.5 |
| Sucrose monolaurate | 2.0 |
| Flavor | 1.0 |
| Sodium saccharinate | 0.1 |
| Water | Balance |
| | <hr/> |
| | 100.0% |

15

20

The above components were blended with 0.05% bovine anti-GTF mother's milk and 0.1% sodium fluoride, 0.005% chlorhexidine gluconate, 0.1% a lytic enzyme, 0.01% a bacteriocin, 0.001% a protease, 0.1% GTF inhibitor-B or 0.25% a dextranase.

25

EXAMPLE 5 Toothpaste

| | | |
|---|---------------------------------------|--------------|
| | Calcium secondary phosphate dihydrate | 50.0% |
| | Glycerol | 20.0 |
| | Sodium carboxymethylcellulose | 2.0 |
| 5 | Sodium lauryl sulfate | 2.0 |
| | Flavor | 1.0 |
| | Sodium saccharinate | 0.1 |
| | Water | Balance |
| | | <hr/> 100.0% |

10 The above components were blended with 0.1% equine anti-protein serum and 0.1% stannous fluoride, 0.01% chlorhexidine gluconate, 0.05% a lytic enzyme, 0.01% a bacteriocin, 0.001% a protease, 0.1% a GTF inhibitor or 0.25% a dextranase.

15 EXAMPLE 6 Toothpaste

| | | |
|----|---------------------------------------|--------------|
| | Calcium secondary phosphate dihydrate | 30.0% |
| | Glycerol | 30.0 |
| | Sorbitol | 20.0 |
| | Sodium carboxymethylcellulose | 1.0 |
| 20 | Sodium lauryl sulfate | 2.0 |
| | Flavor | 1.0 |
| | Sodium saccharinate | 0.1 |
| | Ethanol | 2.0 |
| | Water | Balance |
| 25 | | <hr/> 100.0% |

The above components were blended with 0.1% sheep anti-protein serum and 0.1% stannous fluoride, 0.01%

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chlorhexidine gluconate, 0.1% a lytic enzyme, 0.01% a bacteriocin, 0.0001% a protease, 0.1% GTF inhibitor-A or 0.17% (2000 units/g) a dextranase.

EXAMPLE 7 Toothpowder

| | | |
|----|---------------------------------------|---------|
| 5 | Calcium secondary phosphate dihydrate | 50.0% |
| | Calcium carbonate | 30.0 |
| | Glycerol | 10.0 |
| | α -olefin sulfonate | 1.0 |
| | Flavor | 1.0 |
| 10 | Sodium saccharinate | 0.1 |
| | Dextran | 0.5 |
| | Water | Balance |
| | | 100.0% |

The above components were blended with 0.1% sheep anti-fibrous-substance serum and 0.1% sodium monofluorophosphate and 0.1% sodium fluoride, 0.01% chlorhexidine gluconate, 0.05% a lytic enzyme, 0.001% a bacteriocin, 0.0001% a protease, 0.1% GTF inhibitor or 0.17% a dextranase.

20 EXAMPLE 8 Liquid Dentifrice

| | | |
|----|---------------------|-------|
| | Sodium polyacrylate | 50.0% |
| | Glycerol | 30.0 |
| | Flavor | 0.9 |
| | Sodium saccharinate | 0.1 |
| 25 | Ethanol | 3.0 |
| | Linolic acid | 0.05 |

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| Water | Balance |
|-------|---------|
| | 100.0% |

The above components were blended with 0.01% or 0.02% goat anti-GTF mother's milk and 0.01% or 0.02% goat anti-protein mother's milk and 0.02% sodium fluoride, 0.05% chlorhexidine gluconate, 0.05% a lytic enzyme, 0.001% a bacteriocin, 0.002% a protease, 0.02% GTF inhibitor-A or 0.25% a dextranase.

EXAMPLE 9 Mouthwash

| | | |
|----|---------------------|---------|
| 10 | Ethanol | 20.0% |
| | Flavor | 1.0 |
| | Sodium saccharinate | 0.05 |
| | Sucrose monolaurate | 0.3 |
| | Water | Balance |
| 15 | | 100.0% |

The above components were blended with 0.1% goat anti-GTF serum and 0.1% sodium monofluorophosphate and 0.01% stannous fluoride, 0.01% chlorhexidine gluconate, 0.05% a lytic enzyme, 0.001% a bacteriocin, 0.01% a protease, 0.01% GTF inhibitor-B or 0.25% a dextranase.

EXAMPLE 10 Mouthwash (tablet)

| | | |
|----|----------------------------|-------|
| | Sodium hydrogencarbonate | 54.0% |
| | Sodium secondary phosphate | 10.0 |
| | Polyethylene glycol | 3.0 |
| 25 | Citric acid | 17.0 |
| | Sodium sulfate (anhydrous) | 13.6 |
| | Flavor | 2.0 |

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| | |
|------------|--------|
| Oleic acid | 0.1 |
| | 100.0% |

The above components were blended with 0.1% rabbit anti-GTF serum and 0.1% sodium monofluorophosphate and 0.05% sodium fluoride, 0.05% chlorhexidine gluconate, 0.05% a lytic enzyme, 0.01% a bacteriocin, 0.005% a protease, 0.05% GTF inhibitor-A or 0.25% a dextranase.

The tablet is used by dissolving 0.5 g of the tablet into 50 ml of water.

10 EXAMPLE 11 Gingival Massage Cream

| | | |
|----|--------------------------|---------|
| | White petrolatum | 8.0 |
| | Propylene glycol | 4.0 |
| | Stearyl alcohol | 8.0 |
| | Polyethylene glycol 4000 | 25.0 |
| 15 | Polyethylene glycol 400 | 37.0 |
| | Sucrose stearate | 0.5 |
| | Water | Balance |
| | | 100.0% |

The above components were blended with 0.5% bovine anti-fibrous-substance mother's milk and 0.5% sodium fluoride, 0.01% chlorhexidine gluconate, 0.05% a lytic enzyme, 0.01% a bacteriocin, 0.0% a protease, 0.5% GTF inhibitor-A or 0.25% a dextranase.

25 EXAMPLE 12 Chewing Gum

| | | |
|--|-------------------|--------|
| | Gum base | 43.85% |
| | Calcium carbonate | 2.0 |
| | Starch syrup | 15.0 |

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| | | |
|---|-------------------|--------|
| | Sugar | 30.0 |
| | Sucrose palmitate | 1.0 |
| | Fructose | 4.0 |
| | Maltose | 3.0 |
| 5 | Flavor | 1.0 |
| | | <hr/> |
| | | 100.0% |

The above components were blended with 0.1% bovine anti-whole-bacterial~~body~~ mother's milk and 0.1% stannous fluoride, 0.01% chlorhexidine gluconate, 0.1% a lytic enzyme, 0.01% a bacteriocin, 0.001% a protease, 0.1% GTF inhibitor-C or 0.25% a dextranase.

EXAMPLE 13 Troche

| | | |
|----|-------------------|---------|
| | Gum arabic | 6.0 |
| | Grape sugar | 75.0 |
| 15 | Flavor | 0.2 |
| | <i>l</i> -menthol | 0.1 |
| | Spearmint oil | 0.1 |
| | Sodium ascorbate | 0.1 |
| | Water | Balance |
| 20 | | <hr/> |
| | | 100.0% |

The above components were blended with 0.05% or 0.1% goat anti-protein serum and 0.05% sodium fluoride, 0.01% chlorhexidine gluconate, 0.05% a lytic enzyme, 0.01% a bacteriocin, 0.005% a protease, 0.1% GTF inhibitor-C or 0.25% a dextranase.

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EXAMPLE 14 Dental Paste

| | | |
|----|------------------------------|--------------|
| | Polyoxyethylene monostearate | 2.0% |
| | Sorbitan monooleate | 2.0 |
| | Cetyl alcohol | 2.0 |
| 5 | Palmityl alcohol | 3.0 |
| | Propylene glycol | 15.0 |
| | Carboxymethylcellulose | 5.0 |
| | Saccharine | 0.2 |
| | Peppermint oil | 0.5 |
| 10 | Spearmint oil | 0.5 |
| | Lysozyme chloride | 5000 units/g |
| | Water | Balance |
| | | 100.0% |

The above components were blended with 0.05% or 0.1% equine anti-GTF serum and 0.05% sodium fluoride, 0.01% chlorhexidine hydrochloride, 0.05% a lytic enzyme, 0.01% a bacteriocin, 0.005% a protease, 0.1% GTF inhibitor-A or 0.25% a dextranase.

EXAMPLE 15 Dental Paste

| | | |
|----|----------------------------------|------|
| 20 | Glyceryl monolaurate | 3.0% |
| | Oleyl alcohol | 5.0 |
| | Polyethylene glycol | 15.0 |
| | White petrolatum | 3.0 |
| | N-palmitoyl monosodium glutamate | 0.5 |
| 25 | Hydroxyethylcellulose | 5.0 |
| | Tocopheryl acetate | 0.1 |
| | Sodium saccharinate | 0.2 |

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| | | |
|---|-------------------------|---------|
| | Japanese peppermint oil | 0.7 |
| | Carvone | 0.5 |
| | Anethole | 0.3 |
| | Eugenol | 0.1 |
| 5 | Water | Balance |
| | | <hr/> |
| | | 100.0% |

The above components were blended with 0.025% or 0.05% rabbit anti-fibrous-substance serum and 0.05% sodium fluoride, 0.01% chlorhexidine hydrochloride, 0.05% a lytic enzyme, 0.001% a bacteriocin, 0.0025% a protease, 0.05% GTF inhibitor-B or 0.25% a dextranase.

EXAMPLE 16 Ice-cream

| | | |
|----|----------------------------|--------|
| | Cream (fat content, 50%) | 16.84% |
| | Milk (fat content, 3.7%) * | 42.65 |
| 15 | Defatted evaporated milk | 24.24 |
| | Sugar | 11.25 |
| | Corn syrup | 4.65 |
| | Stabilizer | 0.35 |
| | | <hr/> |
| | | 100.0% |

* : Containing 0.5% bovine anti-fibrous-substance mother's milk

The above components were blended with 0.05% a lytic enzyme or 0.05% a bacteriocin.

EXAMPLE 17 Ice-cream

| | | |
|----|----------------------------|--------|
| 25 | Cream (fat content, 59%) | 16.84% |
| | Milk (fat content, 3.7%) * | 42.65 |
| | Defatted evaporated milk | 24.24 |

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| | |
|------------|---------|
| Sugar | 11.25 |
| Corn syrup | 4.65 |
| Stabilizer | 0.35 |
| | <hr/> |
| | 100.00% |

- 5 * : Containing 3% bovine anti-fibrous-substance
 mother's milk

The above components were blended with 0.001%
a protease, 0.002% GTF inhibitor-C or 0.021% (250 units/g)
a dextranase.

10 EXAMPLE 18 Ice-cream

| | |
|-----------------------------|---------|
| Cream (fat content, 40%) | 31.54% |
| Milk (fat content, 3.7%) ** | 37.16 |
| Defatted evaporated milk | 15.08 |
| Sugar | 11.25 |
| 15 Corn syrup | 4.67 |
| Stabilizer | 0.30 |
| | <hr/> |
| | 100.00% |

** : containing 5% bovine anti-protein mother's milk.

- 20 The above components were blended with 0.05% a
 lytic enzyme, 0.05% a bacteriocin, 0.001% a protease, 0.1%
 GTF inhibitor-A or 0.42% (5000 units/g) a dextranase.

CLAIMS :

1. A caries-preventive composition comprising
an antibody obtained by immunizing a mammal with
at least one antigen selected from the group consisting
of Streptococcus mutans, its cell-wall fraction, fibrous
substance fraction, glucosyltransferase fraction and protein
antigen fraction, and
a synergist selected from the group consisting
of fluorine compounds, chlorhexidine and its salts, lytic
enzymes, bacteriocins, glucosyltransferase inhibitors,
proteases and dextranases.
2. The composition as claimed in claim 1, wherein
Streptococcus mutans is one belonging to the serotype C
separated from human mouth.
3. The composition as claimed in claim 1 or 2, wherein
the antibody is obtained from equine antiserum.
4. The composition as claimed in claim 1 or 2, wherein
the antibody is obtained from bovine antiserum or milk.
5. The composition as claimed in any one of claims 1 to
4, wherein the antibody is prepared from the precipitate
obtained by saturating the antiserum or milk with ammonium
sulfate at the level of not more than 40%.

6. The composition as claimed in any preceding claim, wherein the blending amount of the antibody is in the range of 0.0002 to 10% by weight of the composition.
7. The composition as claimed in any preceding claim,
5 wherein the synergist comprises a fluorine compound selected from the group consisting of monofluorophosphates, alkali-metal fluorides and fluorides containing stannous tin.
8. The composition as claimed in claim 7, wherein the
10 of sodium monofluorophosphate, sodium fluoride, and stannous fluoride.
9. The composition as claimed in any one of claims 1, 7 and 8, wherein the synergist comprises a fluorine compound and the blending amount of the fluoride compound is in
15 the range of 0.0001 to 0.1% by weight of the composition as fluorine.
10. The composition as claimed in any one of claims 1 to 6, wherein the synergist comprises a chlorhexidine salt selected from the group consisting of chlorhexidine
20 hydrochloride and chlorhexidine gluconate.
11. The composition as claimed in claim 1 or claim 10, wherein the synergist comprises chlorhexidine or a chlorhexidine salt and the blending amount of chlorhexidine or its salt is in the range of 0.1 to 1000 ppm.

12. The composition as claimed in any one of claims 1 to 6, including a synergist selected from the group consisting of lytic enzymes, bacteriocins, glucosyltransferase inhibitors, proteases and dextranases and the blending
5 amount thereof is in the range of 0.0001 to 10% by weight of the composition.

13. The composition as claimed in any preceding claim wherein the composition is prepared as a dentifrice, a mouthwash, an oral paste or a gingival massage cream.

10 14. The composition as claimed in any one of claims 10 to 12, wherein the composition is prepared as a troche or a chewing gum.

15. The composition as claimed in claim 12, wherein the composition is prepared as an ice cream.



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EUROPEAN SEARCH REPORT

0140498
Application number

| DOCUMENTS CONSIDERED TO BE RELEVANT | | | EP 84305462.8 |
|---|---|--|--|
| Category | Citation of document with indication, where appropriate, of relevant passages | Relevant to claim | CLASSIFICATION OF THE APPLICATION (Int. Cl. 4) |
| D, Y | GB - A - 1 505 513 (STOLLE RESEARCH AND DEVELOPMENT CO) * Page 1, line 57 - page 2, line 58 * | 1, 2, 4, 13-15 | A 61 K 7/16 A 61 K 39/40 |
| -- | | | |
| Y | WO - A1 - 82/04 396 (THE SECRETARY OF STATE FOR SOCIAL SERVICES IN HER BRITANNIC MAJESTY'S GOVERNMENT) * Claims 5-9; abstract * | 1, 2, 13 | |
| -- | | | |
| Y | GB - A - 2 033 223 (THE SECRETARY OF STATE FOR SOCIAL SERVICES) * Claims 17-21; abstract * | 1, 2, 13 | |
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| Y | GB - A - 2 008 948 (ANIC S.P.A.) * Claims 1-4 * | 1, 12, 13 | A 61 K 7/00 A 61 K 39/00 |
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| Y | CHEMICAL ABSTRACTS, vol. 96, no. 11, March 15, 1982, Columbus, Ohio, USA TOYO JOZO CO. LTD "Glucosyltransferase inhibitor M 5071" page 467, right column, abstract-no. 84 106g & Jpn. Kokai Tokkyo Koho JP 81,103, 193 | 1, 12 | |
| -- | | | |
| The present search report has been drawn up for all claims | | | |
| Place of search VIENNA | | Date of completion of the search 16-11-1984 | Examiner IRMLER |
| CATEGORY OF CITED DOCUMENTS | | | |
| X : particularly relevant if taken alone Y : particularly relevant if combined with another document of the same category A : technological background O : non-written disclosure P : intermediate document | | T : theory or principle underlying the invention E : earlier patent document, but published on, or after the filing date D : document cited in the application L : document cited for other reasons & : member of the same patent family, corresponding document | |

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EUROPEAN SEARCH REPORT

0140498

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|--|---|--|--|
| Category | Citation of document with indication, where appropriate, of relevant passages | Relevant to claim | CLASSIFICATION OF THE APPLICATION (Int. Cl. 4) |
| Y | <p>CHEMICAL ABSTRACTS, vol. 98, no. 3, January 7, 1983, Columbus, Ohio, USA</p> <p>ENDO, AKIRA "Physiologically active mutastein" page 414, right column, abstract-no. 15 482f</p> <p>& Eur. Pat. Appl. EP 59,918</p> <p>----</p> | 1,12 | |
| | | | TECHNICAL FIELDS SEARCHED (Int. Cl. 4) |
| | | | |
| The present search report has been drawn up for all claims | | | |
| Place of search VIENNA | | Date of completion of the search 16-11-1984 | Examiner IRMLER |
| <p>CATEGORY OF CITED DOCUMENTS</p> <p>X : particularly relevant if taken alone</p> <p>Y : particularly relevant if combined with another document of the same category</p> <p>A : technological background</p> <p>O : non-written disclosure</p> <p>P : intermediate document</p> <p>T : theory or principle underlying the invention</p> <p>E : earlier patent document, but published on, or after the filing date</p> <p>D : document cited in the application</p> <p>L : document cited for other reasons</p> <p>& : member of the same patent family, corresponding document</p> | | | |

